

Frontiers in Food, Drug and Natural Sciences (2025), Vol 2: 9-15

An international peer-reviewed online journal

Available online: fd-science.com



Review Article

An overview of some of the packaging methods for fish

Behnam Farjami, Nargess Anoosheh*

Department of Fisheries, Faculty of Natural Resources, University of Tehran, Karaj, POBox: 4111, Iran

ARTICLE INFO

This article was previously published in Persian Journal of Seafood Science and Technology (2015, 1: 36-40). However, the journal's name was changed to Frontiers in Food, Drug and Natural Sciences (FDNS). For the citation purposes and courtesy of the authors, this article is re-published in (FDNS).

ABSTRACT

Food packaging is necessary for storage, suitable distribution and maintaining the quality. Fish has a high nutritional value and their consumption is highly recommended. Per capita consumption of fish is low in Iran. This is due to factors such as lack of access to high quality and fresh fish and supplied of without packaging or improper packaging. Marine products are more susceptible to spoilage than other meat products. Packaging is one of the most common and easy methods to maintain the freshness of fisheries products from the catch to consumption. There are different methods for packaging of fish and its products. Some of the methods such as modified atmosphere packaging (MAP) and vacuum packaging are widely used for fisheries products. MAP has become a popular method for packaging of foods as it can extend the shelf life of food. One of the new methods of seafood packaging is the edible films and coating. Edible films and coatings are thin layers which are used both in surface and in between the various layers of food. In this paper, three common packaging methods of seafood i.e. MAP, vacuum packaging and edible films and coatings are reviewed.

Keywords: Edible coating; Fish; MAP; Packaging; Shelf-life.

^{*} Direct inquiries to author: n_anoosheh@alumni.ut.ac.ir

1. Introduction

Fresh fish may provide a higher quality and nutritious source of protein but this has been facing problems in terms of quality and distribution. Therefore, providing solutions to maintain the quality and systematic distribution of fresh fish is required. In other words, we must find ways to store and transport fresh fish in order to maintain its quality for a longer period and offer it to the remote regions where there is no possibility of farming and production (Mendes & Goncalves, 2008).

Seafood due to their high protein content, fatsoluble vitamins and omega-3 polyunsaturated fatty acids, which are important in human diets, have attracted much attention (Pérez-Alonso et al., 2004). Seafood products are perishable and are usually spoiled faster than other meat products. Flesh of seafood is susceptible to changes more than other meats, after death. This may be due to different composition of seafood and other meats (Stamatis & Arkoudelos, 2007; Hosseini et al., 2013; Gringer et al., 2015). Fish spoilage occurs through biological reactions including autolysis, lipid oxidation and activity of microorganisms (Gobantes et al., 1998). Fish spoilage rate depends on the quality of primary and storage conditions. Factors such as temperature storage, processing and packaging are the most important factors in the shelf-life of seafood (Moini et al., 2009; Alikhani Chamgordani et al., 2024).

Packaging is a protective method that maintains the quality of goods from production to consumption. Packaging protects the product against climatic changes, injuries, damages, humidity, vibration, odors, pressure, microorganisms and insects (Farjamia & Anoosheha, 2015).

Some common packaging method in the seafood industry include packaging in plastic, glass, metal, modified atmosphere packaging, vacuum packaging, packaging with edible coating, etc. Packaging is the simplest method that can be used from the earliest stage (mean catch) to the consumption. Therefore, it seems necessary to investigate the packaging of fish and marine products and the advantages and

disadvantage of these methods. The aim of this study is to review some conventional packaging in aquatic products (Farshbaf Aghajani *et al.*, 2023).

2. Modified atmosphere packaging (MAP)

Modified atmosphere packaging (MAP) is used as a supplement to traditional storage method and will be long term food storage (Arritt et al., 2007). Usually, MAP packaging gases include oxygen (O_2) , carbon dioxide (CO_2) and nitrogen (N_2) . The proportions of each gas are established, the mixture is introduced into the packaging and no further control is carried out during storage (Silliker & Sk, 1980). Combination gas in MAP packaging depends on the type of food product (Velu et al., 2013). MAP packaging has an important role in increasing the shelf life of susceptible foods to spoilage at refrigerator temperature (Parry, 2012). Reason of this is the effect of CO₂ gas to inhibit the growth of spoilage bacteria (Magsood & Benjakul, 2010; Provincial et al., 2010). The role of O2 is inhibition of the of absolute anaerobic bacteria (Rutherford et al., 2007). In food presenting high moisture and/or fat amounts, such as fish, beef and poultry, the excessive absorption of CO₂ may lead to a phenomenon known as "packaging collapse" (Parry, 2012). Increase in dripping is caused by the dissolution of gases on the muscles surface in atmospheres containing high CO₂ levels (>60%), reduced pH and, consequently, low protein water retention ability (Randell et al., 1995; Parry, 2012; Farshbaf Aghajani et al., 2024).

The presence of oxygen may cause oxidative rancidity problems in fish presenting high lipids amounts, promoting the formation of low molecular weight aldehydes, ketones, alcohols and carboxylic acids. Thus, the use of O₂ in modified atmospheres is generally avoided with this kind of fish, in order to minimize such effects. Davies and Slade (1995) stated that there are evidences showing that the use of O₂ reduces the exudation in fish during storage. In addition, other factors such as the characteristics of packaging and fish species are effective on the shelf life of product (Otwell *et al.*, 2008). As a

consequence, high CO2 concentrations promote organoleptic changes as, for example, texture alterations in meat. N2 can be used as an inert gas in smaller proportions than CO2. Nitrogen has low solubility in water and has no taste, color and odor. Nitrogen plays a role in delaying oxidative rancidity and inhibiting aerobic bacterial growth. However, it is not capable in inhibiting anaerobic bacterial growth. The oxygen can also be employed, providing fish does not undergo color alterations (Cann et al., 1983). Although studies have shown that the MAP packaging increases the shelf life of seafood, but given that the composition of fish and storage conditions in different regions and seasons, therefore the combination of these gases and their effects in order to achieve optimum condition are different. Hence, to achieve the best condition for longer storage and delivering safe product to consumers is necessary (Otwell et al., 2008). Özogul et al. (2004) had investigated the effect of MAP (CO₂/N₂: 60/40) on total viable count and histamine forming bacteria count in sardines (Sardina pilchardus) kept at 4°C. The outcome illustrated the lowest total viable count (TVC) in MAP compared to vacuum packaging and air.

3. Vacuum packaging

Vacuum packaging is a suitable way to retard the spoilage of marine products which increases the shelf life and preserving the quality of fish for longer duration. Exhaust of O2 of this package not only will be delaying microbial spoilage, but also delay the non-microbial spoilage of product. Thus this method increases the shelf life of meat products and preserves their quality and freshness during storage (Sahoo & Kumar, 2005). The gaseous atmosphere of the vacuum packaging is reduced, but it is probably altered during storage, thus considered modified due to a 10 to 20% increase in the CO₂ amount produced by microbial activity. This CO2 may inhibit the growth of undesirable microorganisms (Silliker & Sk, 1980). It is reported that, the solubility of produced CO2 in muscles has stabilizing effect on the pH in this packaging. Therefore, it prevents the increase of the volatile nitrogenous compound and produced ammonium by bacteria that effect on the pH (Mendes & Gonçalves,

2008). Farjamia and Anoosheha (2015) investigated the effect of vacuum packaging on the properties of chemical, microbial and sensory of Rutillus frisii kutum. Their results showed that the vacuum packaging increases the shelf life and preserves chemical, microbial and sensory quality of fish fillet on the ice. Özogul et al. (2004) investigated the effect of MAP and vacuum packaging on the chemical, microbial and sensory changes of Sardin pichardus. Their results showed that the bacteria had the fastest growth in stored sardine in combination with normal air and the lowest count of bacteria occurred in the MAP with gas mixture (40% N₂ and 60% CO₂). Kosak and Toledo (1981) studied the combination of a chlorine solution (1000 mg/mL free chlorine) with vacuum polyethylene packaging for mullet (Mugil cephalus) stored at -2 °C. All treatments were organoleptically acceptable up to 14 days of storage.

4. Edible films and coatings

Nowadays, pollution of synthetic polymers has focused public attention on the use of biodegradable material. During the past two decades, the application of biodegradable material from the protein and carbohydrates has been widely studied. These macromolecules can potentially be used as an alternative to synthetic petroleum polymers from derivatives (Ghanbarzadeh al.. Edible et 2007). biodegradable packaging are divided into two categories of edible films and coatings (Ghanbarzadeh & Oromiehi, 2008). Edible films are produced as a thin layer before use in food packaging and used as synthetic polymers for packaging (Ghanbarzadeh et al., 2006). Films can be created as a wrapper, capsule and bags. These products are molded thick. Unlike films, edible coatings are formed on the food. Therefore, the coating is part of the product and remains on the product in using time. This can be done by methods such as polish, spray and immersion (Ghanbarzade et al., 2007). Various edible coatings of polysaccharides, proteins and fats can increase the shelf life of food products and play a key role as barrier against the transfer of moisture, gases and dissolved substances. These coatings due to their biodegradable

characteristic have a lot of popularity among consumers and can be used as an alternative to the existing plastic material in the packaging industry (Wang et al., 2010). Numerous studies have been conducted on the application of edible films and coatings for packaging and storage of aquatic products. (Mohan et al., 2012) investigated the effect of chitosan film on the quality of Sardinella longiceps. Their results showed that the sensory quality was maintained during storage. Also, the microbial growth, oxidation of lipids, total volatile nitrogen bases (TVB-N), drip and water-holding capacity were reduced. Soares et al. (2013) studied the effect of chitosan coating and glazing on the salmon fish. Results showed that this coating controlled the lipid oxidation and microbial growth. Also, chitosan and glazing increase the shelf life and decrease drip. Chitosan has antioxidants (López-Caballero et al., 2005), antimicrobial properties (Jeon et al., 2002) and ability to connect (Shahidi et al., 1999). The antioxidant effect of chitosan is due to amine groups of chitosan that constitute a stable compound with volatile aldehydes such as malondialdehyde (Weist & Karel, 1992; Shahidi et al., 1999). Also, the chitosan can reduce lipid oxidation by chelating the Fe2+ ions present in fish protein. Therefore it eliminates the peroxidation activity by converting to the Fe3+ (Jeon et al., 2002). Several factors affect the antimicrobial action of chitosan (Cuero, 1999). Some of these factors are the reaction between the molecules of positively charged of chitosan and negatively cell membranes of microbes (Shahidi et al., 1999). Also, chitosan is a barrier against O₂ (Jeon et al., 2002). In addition, the antimicrobial effect of chitosan related to the elimination of the lipopolysaccharide of outer membrane of Gram negative bacteria (Nikaido, 1996; Helander et al., 2001). Rodriguez-Turienzo et al. (2011) examined the whey protein concentrate on the Salmo salar. This coating reduced the lipid oxidation, microbial growth and drip and preserved the sensory quality of fish. Franssen et al. (2004) indicated that the whey protein has excellent protective properties against O2. In study of Hasanzati Rostami et al. (2010), reducing the amount of peroxide in Kilka covered with the whey may be due to the

protective properties of whey against O2. Khanehdan (2011) investigated the effect of sodium alginate film on the Kilka. The results showed that total number of bacteria of fish reduced. Alginate has antioxidant properties, antimicrobial and protection against penetration of water vapor. Song et al. (2011) reported that alginate plays the role of a barrier against O2 transfer and prevents the growth of aerobic bacteria. It also delays the lipids oxidation. Alginate has gel-forming properties. Thus, it produces a semi-permeable layer and traps the microorganisms, effectively. Khanehdan (2011) concluded that should be used in the lipid-based materials in combination with sodium alginate film to prevent the loss of moisture in stored Kilka in the freezer (Dehghani et al., 2018).

5. Conclusions and perspectives

Consumer demand for MAP has increased due to its technical opportunity with a significant effect in extending the shelf life of fishery products, providing good quality food products, and avoiding or lessening usage of chemical preservatives. The use of MAP can delay the growth of many spoilage microorganisms and inhibit their toxin production. Vacuum packaging effectively controls the production of volatile nitrogenous base but cannot control the lipid oxidation during storage in ice. According to previously researches, vacuum packaging in comparison with packaging in normal conditions can maintain the quality and freshness of fish in longer term (Moini et al., 2009).

Biopolymers such polysaccharides, as proteins and lipids are used to produce edible films and coatings. Some of the properties of edible films and coatings include preventing migration of moisture and exit of volatile compounds of food, delaying of dehydration, a barrier against microbial growth, lipid oxidation and drip and protecting the quality of marine products against physical damage. There are several materials that are used to produce edible films and coatings but, more studies are needed to improve the production methods of films and their applications in aquatic food packaging (Hosseini et al., 2009; Bahmani et al., 2011; Dehghani et al., 2019).

Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

6. References

- Alikhani Chamgordani, P., Soltani Firouz, M., Omid, M., Hadidi, N., & Farshbaf Aghajani, P. (2024). Dual-stage ultrasound in deep frying of potato chips; effects on the oil absorption and the quality of fried chips. *Ultrasonics Sonochemistry*, 103, 106779.
- Arritt, F. M., Eifert, J. D., Jahncke, M. L., Pierson, M. D., & Williams, R. C. (2007). Effects of modified atmosphere packaging on toxin production by *Clostridium botulinum* in raw aquacultured summer flounder fillets (*Paralichthys dentatus*). *Journal of Food Protection*, 70(5), 1159–1164.
- Bahmani, Z. A., Rezai, M., Hosseini, S. V., Regenstein, J. M., Böhme, K., Alishahi, A., & Yadollahi, F. (2011). Chilled storage of golden gray mullet (*Liza aurata*). *LWT Food Science and Technology*, 44(9), 1894–1900.
- Cann, D. C., Smith, G. L., & Houston, N. C. (1983). Further studies on marine fish stored under modified atmosphere packaging. Scotland: Torry Research Station.
- Cuero, R. G. (1999). Antimicrobial action of exogenous chitosan. *Exs*, *87*, 315–333.
- Davies, A. R., & Slade, A. (1995). Fate of *Aeromonas* and *Yersinia* on modified-atmosphere-packaged (MAP) cod and trout. *Letters in Applied Microbiology*, 21(6), 354–358.
- Dehghani, S., Hosseini, S. V., & Regenstein, J. M. (2018). Edible films and coatings in seafood preservation: A review. *Food Chemistry*, 240, 505–513.
- Dehghani, S., Peighambardoust, S. H., Peighambardoust, S. J., Hosseini, S. V., & Regenstein, J. M. (2019). Improved mechanical and antibacterial properties of active LDPE films prepared with combination of Ag, ZnO and CuO nanoparticles. Food Packaging and Shelf Life, 22, 100391.
- Farjamia, B., & Anoosheha, N. (2015). An overview of some of the packaging methods of fish. *Journal of Seafood Science and*

- Technology, 1, 1-5.
- Farshbaf Aghajani, P., Soltani Firouz, M., & Alikhani Chamgordani, P. (2023). The improvement of freezing time and functional quality of frozen mushrooms by application of probe-type power ultrasound. *Ultrasonics Sonochemistry*, 100, 106637.
- Farshbaf Aghajani, P., Soltani Firouz, M., & Bohlol, P. (2024). Revolutionizing mushroom identification: Improving efficiency with ultrasound-assisted frozen sample analysis and deep learning techniques. *Journal of Agriculture and Food Research*, 15, 100946.
- Franssen, L. R., Rumsey, T. R., & Krochta, J. M. (2004). Whey protein film composition effects on potassium sorbate and natamycin diffusion. *Journal of Food Science*, 69(5), C347–C350.
- Ghanbarzadeh, B., Musavi, M., Oromiehie, A. R., Rezayi, K., Rad, E. R., & Milani, J. (2007). Effect of plasticizing sugars on water vapor permeability, surface energy and microstructure properties of zein films. *LWT-Food Science and Technology*, 40(7), 1191–1197.
- Ghanbarzadeh, B., & Oromiehi, A. R. (2008). Biodegradable biocomposite films based on whey protein and zein: Barrier, mechanical properties and AFM analysis. *International Journal of Biological Macromolecules*, 43(2), 209–215.
- Ghanbarzadeh, B., Ouroumiehei, A. A., Mousavi, M., Razmi, E., & Milani, J. (2006). Effect of polyolic plasticizers on rheological and thermal properties of zein resins. *Iranian Polymer Journal*, *15*(10), 779–787.
- Gobantes, I., Choubert, G., & Gómez, R. (1998). Quality of pigmented (astaxanthin and canthaxanthin) rainbow trout (*Oncorhynchus mykiss*) fillets stored under vacuum packaging during chilled storage. *Journal of Agricultural and Food Chemistry*, 46(10), 4358–4362.
- Gringer, N., Hosseini, S. V., Svendsen, T., Undeland, I., Christensen, M. L., & Baron, C.
 P. (2015). Recovery of biomolecules from marinated herring (*Clupea harengus*) brine using ultrafiltration through ceramic membranes. *LWT Food Science and Technology*, 63(1), 423–429.

- Hasanzati Rostami, A., Motallebi, A. A.,
 Khanipour, A. A., Soltani, M., & Khanedan, N.
 (2010). Effect of whey protein coating on physic-chemical properties of gutted Kilka during frozen storage. *Iranian Journal of Fisheries Sciences*, 9(3), 412–421.
- Helander, I. M., Nurmiaho-Lassila, E. L., Ahvenainen, R., Rhoades, J., & Roller, S. (2001). Chitosan disrupts the barrier properties of the outer membrane of Gramnegative bacteria. *International Journal of Food Microbiology*, 71(2–3), 235–244.
- Hosseini, S. V., Arlindo, S., Böhme, K., Fernández-No, C., Calo-Mata, P., & Barros-Velázquez, J. (2009). Molecular and probiotic characterization of bacteriocin-producing *Enterococcus faecium* strains isolated from nonfermented animal foods. *Journal of Applied Microbiology*, 107(4), 1392–1403.
- Hosseini, S. V., Sobhanardakani, S., Tahergorabi, R., & Delfieh, P. (2013). Selected heavy metals analysis of Persian sturgeon's (*Acipenser persicus*) caviar from southern Caspian Sea. *Biological Trace Element Research*, 154, 357–362.
- Jeon, Y.-J., Kamil, J. Y. V. A., & Shahidi, F. (2002). Chitosan as an edible invisible film for quality preservation of herring and Atlantic cod. *Journal of Agricultural and Food Chemistry*, 50(18), 5167–5178.
- Khanehdan, N., Motalebi, A. A., Khanipour, A. A., Koochekian Sabour, A., Seifzadeh, M., & Hasanzati Rostami, A. (2011). Effects of different concentrations of sodium alginate as an edible film on chemical changes of dressed Kilka during frozen storage. *Iranian Journal of Fisheries Sciences*, 10(4), 654–662.
- López-Caballero, M. E., Gómez-Guillén, M. C., Pérez-Mateos, M., & Montero, P. (2005). A chitosan–gelatin blend as a coating for fish patties. *Food Hydrocolloids*, 19(2), 303–311.
- Maqsood, S., & Benjakul, S. (2010). Synergistic effect of tannic acid and modified atmospheric packaging on the prevention of lipid oxidation and quality losses of refrigerated striped catfish slices. *Food Chemistry*, 121(1), 29–38.
- Mendes, R., & Gonçalves, A. (2008). Effect of soluble CO2 stabilisation and vacuum packaging in the shelf life of farmed sea bream and sea bass fillets. *International Journal of Food Science & Technology*, 43(9), 1678–

1687.

- Mohan, C. O., Ravishankar, C. N., Lalitha, K. V., & Gopal, T. K. S. (2012). Effect of chitosan edible coating on the quality of double filleted Indian oil sardine (*Sardinella longiceps*) during chilled storage. *Food Hydrocolloids*, 26(1), 167–174.
- Moini, S., Tahergorabi, R., Hosseini, S. V., Rabbani, M., Tahergorabi, Z., Feás, X., & Aflaki, F. (2009). Effect of gamma radiation on the quality and shelf life of refrigerated rainbow trout (*Oncorhynchus mykiss*) fillets. *Journal of Food Protection*, 72(7), 1419–1426.
- Nikaido, H. (1996). Outer membrane. In *Escherichia coli and Salmonella typhimurium: Cellular and molecular biology* (2nd ed., pp. 29–47). Washington, DC: ASM Press.
- Otwell, W. S., Kristinsson, H. G., & Balaban, M. O. (2008). Modified atmospheric processing and packaging of fish: Filtered smokes, carbon monoxide, and reduced oxygen packaging. Hoboken, NJ: John Wiley & Sons.
- Özogul, F., Polat, A., & Özogul, Y. (2004). The effects of modified atmosphere packaging and vacuum packaging on chemical, sensory and microbiological changes of sardines (*Sardina pilchardus*). *Food Chemistry*, 85(1), 49–57.
- Parry, R. T. (2012). *Principles and applications* of modified atmosphere packaging of foods. London, UK: Blackie Academic & Professional.
- Pérez-Alonso, F., Aubourg, S. P., Rodríguez, Ó., & Barros-Velázquez, J. (2004). Shelf life extension of Atlantic pomfret (*Brama brama*) fillets by packaging under a vacuum-skin system. *European Food Research and Technology*, 218, 313–317.
- Provincial, L., Gil, M., Guillén, E., Alonso, V., Roncalés, P., & Beltrán, J. A. (2010). Effect of modified atmosphere packaging different CO2 and N2 combinations on chemical, microbiological physical, and of changes fresh sensory sea (Dicentrarchus labrax) fillets. International Journal of Food Science & Technology, 45(9), 1828-1836.
- Randell, K., Ahvenainen, R., & Hattula, T. (1995). Effect of the gas/product ratio and CO2 concentration on the shelf life of MA packed

- fish. *Packaging Technology and Science*, 8(4), 205–218.
- Rodriguez-Turienzo, L., Cobos, A., Moreno, V., Caride, A., Vieites, J. M., & Diaz, O. (2011). Whey protein-based coatings on frozen Atlantic salmon (*Salmo salar*): Influence of the plasticiser and the moment of coating on quality preservation. *Food Chemistry*, 128(1), 187–194.
- Rutherford, T. J., Marshall, D. L., Andrews, L. S., Coggins, P. C., Schilling, M. W., & Gerard, P. (2007). Combined effect of packaging atmosphere and storage temperature on growth of *Listeria monocytogenes* on readyto-eat shrimp. *Food Microbiology*, 24(7–8), 703–710.
- Sahoo, J., & Kumar, N. (2005). Quality of vacuum packaged muscle foods stored under frozen conditions: A review. *Journal of Food Science and Technology*, *42*(3), 209–213.
- Shahidi, F., Arachchi, J. K. V., & Jeon, Y.-J. (1999). Food applications of chitin and chitosans. *Trends in Food Science & Technology*, 10(2), 37–51.
- Silliker, J. H., & Sk, W. (1980). Microbiological safety considerations in controlled atmosphere storage of meats. *Food Technology*, *34*(3), 59–63.
- Soares, N. M., Mendes, T. S., & Vicente, A. A. (2013). Effect of chitosan-based solutions applied as edible coatings and water glazing on frozen salmon preservation: A pilot-scale study. *Journal of Food Engineering*, 119(2), 316–323.

- Song, Y., Liu, L., Shen, H., You, J., & Luo, Y. (2011). Effect of sodium alginate-based edible coating containing different antioxidants on quality and shelf life of refrigerated bream (Megalobrama amblycephala). Food Control, 22(3–4), 608–615.
- Stamatis, N., & Arkoudelos, J. S. (2007). Effect of modified atmosphere and vacuum packaging on microbial, chemical and sensory quality indicators of fresh, filleted *Sardina pilchardus* at 3 °C. *Journal of the Science of Food and Agriculture*, 87(6), 1164–1171.
- Velu, S., Abu Bakar, F., Mahyudin, N. A., Saari, N., & Zaman, M. Z. (2013). Effect of modified atmosphere packaging on microbial flora changes in fishery products. *International Food Research Journal*, 20(1), 1–10.
- Wang, L., Auty, M. A. E., & Kerry, J. P. (2010). Physical assessment of composite biodegradable films manufactured using whey protein isolate, gelatin and sodium alginate. *Journal of Food Engineering*, 96(2), 199–207.
- Weist, J. L., & Karel, M. (1992). Development of a fluorescence sensor to monitor lipid oxidation. 1. Fluorescence spectra of chitosan powder and polyamide powder after exposure to volatile lipid oxidation products. *Journal of Agricultural and Food Chemistry*, 40(7), 1158–1162.